

# FORMULATION AND EVALUATION OF NANOFORMULATIONS OF BICALUTAMIDE

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## Abstract:

Bicalutamide (BIC) is a non-steroidal anti-androgen used for monotherapy in treating prostate cancer. The nanoparticles of bicalutamide were prepared by solvent evaporation method and evaluated for FT-IR, DSC, X-ray diffraction in vitro dissolution and in vivo diffusion studies. The spectra of unfunctionalized MSNs were comparatively simple and major peaks could be easily assigned. A characteristic peak attributed to BIC could be seen. The melting point of BIC was found to be 192.54 °C denoted by a sharp endothermic peak. The complete entrapment of drug into mesoporous carriers was confirmed by absence of any such peak in the thermogram. This pattern was well preserved and even after functionalization and drug loading. BIC release from MCM-41 was highest followed by marketed formulation. It causes dose dependent reduction in the Prostate specific antigen (PSA) levels. It has the potential of working wonders at all the stages of prostate cancer disease continuum. Thus, the article reveals the application of bare and functionalized MSNs as oral as well as intravenous targeted delivery agents for BIC in cancer therapy have been proposed in depth along with their biosafety aspects and efficacy.

**Keywords:** Nanoformulation, FTIR, DSC, In vitro dissolution, XRD

## Introduction:

Bicalutamide (BIC) is a non-steroidal anti-androgen used for monotherapy in treating prostate cancer (1). It causes dose dependent reduction in the Prostate specific antigen (PSA) levels. It has the potential of working wonders at all the stages of prostate cancer disease continuum (2). BIC binds to cytoplasmic androgenic receptors and competitively inhibits the androgenic action by producing distortion of the co-activator binding site, thereby stopping the initiation of gene transcription. It causes some central androgen blockade and not much affects the testosterone and LH levels. It is also approved for combination therapy along

with LHRH analogue for treating metastatic prostate cancer. It possesses long  $t_{1/2}$  and undergoes an extensive metabolism in liver (3). Some of the major challenges currently faced by the current cancer treatment include the solubility and permeability limitations faced by most of the new chemical entities and anticancer agents. Another important challenge is reducing the side effects of the current chemotherapeutic treatment by aiming for tumour targeted therapy for cancer. The most recent advancement being "theranostics" in which simultaneous diagnosis and the therapeutic treatment is possible by incorporating both the agents in a single carrier. BIC comes under Biopharmaceutical classification system (BCS) Class II and suffers from solubility limitations, leading to dissolution and bioavailability issues. Thus, formulating an novel drug delivery system for this drug could open new avenues for increasing its effectiveness in prostate cancer treatment. To achieve this purpose a mesoporous silica nano delivery system was designed using MCM-41 carriers and thereafter functionalizing with ligands and surface moieties which help in achieving a targeted mesoporous system for the drug BIC.

The drug delivery systems were designed based on the pH stimuli based and another was based on targeting the receptors overexpressed in cancer like folate receptors. PAA polymer was used as a pH responsive material and coated onto the surface of MCM-41 MSNs via an aminated intermediate layer. This was based on the fact that environment of cancer cells is more acidic than that of the healthy cells. Also, certain receptors are found to be overexpressed in cancer cells like folate receptors. Folic acid was used as a ligand to target the overexpressed folate receptors in cancer.

Thus, the article reveals the application of bare and functionalized MSNs as oral as well as intravenous targeted delivery agents for BIC in cancer therapy have been proposed in depth along with their biosafety aspects and efficacy.

The facile synthesis and functionalization strategy was adopted. Dissolution and Caco-2 Permeability study was

performed and Pharmacokinetic data and biodistribution analysis was noted and cytotoxicity study was done on LNCaP and PC-3 prostate cancer cell lines. The cell death mechanism, cell killing efficiency and cellular uptake was determined. A thorough histological examination was done to adjudge their preliminary biosafety.

#### Materials and Methods:

##### Materials

Bicalutamide (BIC) was obtained as a sample gratis from Intas Pharmaceuticals Ltd, (Ahmedabad, Gujarat, India). Other Chemicals required for the research study were purchased from S.D. Fine Chem Ltd, Mumbai.

##### Methods:

##### Synthesis of bare and functionalized MSNs

Aminated MCM-41 NPs were obtained by reaction of MCM-41 NPs with APTES as per a reported method in the literature (2). Typically, 250 mg of MCM-41 was accurately weighed and transferred to a round bottom flask (RBF). The MCM-41 MSNs were dispersed in 30 mL toluene. Thereafter, 3.43 mL of APTES was added. The reaction was kept under vigorous stirring at 70°C for 12h. Thereafter, RBF was allowed to cool slowly at RT. The reaction mixture obtained was filtered and washed with methanol. The product obtained was dried and labelled as MCM-41-A. Successful functionalization with APTES laid a strong foundation for further functionalization with Polyacrylic acid (PAA). Differential scanning calorimetry (DSC) analysis: Melting point and loading were determined by DSC Shimadzu-TA60 thermal analyzer equipped with the TA60

WS software. The heating rate was kept at 10°C/min. The existence of BIC and BIC on silica matrix in amorphous or crystalline form and complete encapsulation was further confirmed by DSC. Absence of any melting point depression is indicative of presence of drug in non-crystalline state in pores (8). Melting points of BIC and BIC were determined individually. DSC analysis of BIC-MCM-41, BIC-MCM-41-A, BIC-PAA-MSN, BIC-FA-MSN, BIC-MCM-41, BIC-MCM-41-A, BIC-

PAA-MSN, BIC-FA-MSN was done and absence of any crystalline sharp drug peaks was ensured.

##### X-Ray diffraction analysis (XRD)

PA analytical model equipped with Cu K radiation beam operating at 40 kV and 40 mA was used to determine low

angle X-Ray powder diffraction (XRD) pattern. The structure of pores was ascertained by low-angle XRD measurements. The spectra was an indication of the intactness of the mesoporous structure. XRD measurements were taken for MCM-41, BIC-MCM-41, MCM-41-A, BIC-MCM-41-A, PAA-MSN, BIC-PAA-MSN, FA-MSN and BIC-MCM-41, BIC-MCM-41-A, BIC-PAA-MSN and BIC-FA-MSN.

Morphological characterization was performed using scanning electron microscopy (SEM) (FEI-Quanta 200 operating at 20 kv) (Thermo Scientific, USA). The samples were coated with gold to make them conducting before imaging. The images were resolved over a photographic film. SEM analysis was done to determine morphological and internal structure of bare MCM-41 and functionalized PAA-MSN and FA-MSNs respectively.

##### Dissolution Kinetics study

The % cumulative release data obtained from dissolution study was fitted to various kinetic models and the best fit was determined based on the R<sup>2</sup> value, AIC criterion and Model selection criterion.

##### In vitro diffusion study

Phosphate buffer saline (PBS) media of different pH like 5.5, 6.8 and 7.4 was used to determine the in vitro release behaviour of BIC. A dialysis tube having cut-off Molecular weight (Mw) of 7000 g/mol was used to fill the suspension of BIC, BIC-MCM-41, BIC-MCM-41-A, BIC-PAA-MSNs and BIC-FA-MSN. Continuous magnetic stirring was provided and sink conditions were maintained properly by replacing withdrawn samples immediately with fresh PBS of respective pH. The withdrawn samples were analysed by measuring the fluorescence intensity by Spectrofluorometer keeping affixed excitation wavelength of 260 nm and measuring.

##### Zeta potential and size determination:

Particle Z-average size and charge were measured using dynamic light scattering (DLS) and electrophoretic mobility measurement respectively, using the Malvern Zetasizer Nano ZS (Malvern instruments, Malvern, UK). Zeta potential indicates the surface residual charge of the particles. The zeta potential measurements were done for MCM-41, MCM-41-A, PAA-MSN, FA-MSN and drug loaded MCM-41, MCM-41-A, PAA-MSN and FA-MSN.

##### In vitro release study:

The drug release pattern was determined and cumulative drug release was calculated for all the mesoporous formulations of BIC. For formulations to be administered by oral route, dissolution study was performed. Additionally simulated and biorelevant media was used to study the effect of enzymes and food on drug release. For parenteral formulations, in vitro diffusion study was performed at different pH values using PBS medium.

#### In vitro dissolution study

Dissolution study was performed using Veego USP type II dissolution apparatus in 900 mL dissolution media at 50 rpm maintaining temperature of dissolution medium at  $37 \pm 0.5^\circ\text{C}$ . The in vitro release study was performed for plain BIC (API), MF, BIC-MCM-41 and BIC-MCM-41-A in acetate media (pH 4.6), simulated gastric fluid (SGF) (pH 1.2) and simulated intestinal fluid (SIF) (pH 6.8) containing pepsin and pancreatin respectively. Enzyme free SGF (pH 1.2) and SIF (pH 6.8) were also taken to study the presence of any interaction between hard gelatin capsule shell and amine group of MCM-41-A and whether it has any effect on the release of BIC. The drug release pattern was also studied in the presence and absence of food as well. Hence, the fed and fasted state simulated gastric and intestinal media were prepared (FaSSGF, FeSSGF, FaSSIF, FeSSIF) for this purpose. The composition of all media is summarized in Table 6.1 (23, 24). The powder was filled in the hard gelatin capsule shell prior to dissolution study. 5 mL aliquots were withdrawn at 5, 10, 15, 20, 30, 45, 60, 90, 120, 180, 240, 320 and 360 min intervals. The withdrawn samples were filtered through 0.45  $\mu\text{m}$  PVDF filter membrane and analysed by UV spectrophotometer at 285 nm. Sink conditions were maintained throughout the study by adding an equivalent amount of fresh medium as that of withdrawn sample.

#### Dissolution Kinetics study

For the purpose of quantifying the differences in the release profiles of BIC formulations. The drug release data from both MCM-41 and MCM-41-A nanoparticles was fitted to various kinetic models and the best fit was determined. Different parameters by fitting the experimental data to different release models were calculated. Criteria for judgement on **best model included lowest AIC (Akaike information criterion), highest MSC (Model selection criterion) and Regression values (R<sup>2</sup>) (25). The various release models to which dissolution data were fitted include zero order, first order, Higuchi, Weibull, Hixon-Crowell and Korsmeyer-peppas model (26).**

#### In vitro diffusion study

In the in vitro drug release study, a suspension of BIC, and BIC-MCM-41, BIC-MCM-41-A, BIC-PAA-MSN and BIC-FA-MSN was filled into a dialysis tube (cut-off Molecular weight (Mw) = 7000 g/mol for PAA-MSNs) and (cut-off Mw = 3500 for FA-MSNs). Further, the bag containing filled dispersion was then immersed into 100 mL

#### In vitro dissolution study

Veego USP type II paddle apparatus was used to perform the dissolution study in 1000 mL dissolution media at 50 rpm maintaining  $37 \pm 0.5^\circ\text{C}$  temperature. The in vitro dissolution study was performed for BIC, Marketed Formulation (MF), BIC-MCM-41 and BIC-MCM-41-A in various media viz water (with addition of 0.5% SLS), FaSSGF, FeSSGF, FaSSIF, and FeSSIF (Fast and Fed state biorelevant media) to study the drug release pattern from non-functionalized

#### Results and Discussion

##### Fourier Transform Infra Red (FT-IR) spectroscopy studies

FT-IR spectra (Figure. 1) provided proof of the manner in which successful synthesis and functionalization of MCM-41 mesoporous silica nanoparticles proceeded. The spectra of unfunctionalized MSNs was comparatively simple and major peaks could be easily assigned. In a characteristic peak attributed to BIC could be seen. The cyanide and amide carbonyl groups were denoted by peaks obtained at  $2227\text{cm}^{-1}$ ,  $1685\text{cm}^{-1}$  and  $3341\text{cm}^{-1}$  respectively.

Aromatic stretching due to phenyl group was observed at  $708$  and  $833\text{cm}^{-1}$ . Characteristic cyano and carbonyl group peaks were seen at wavenumbers  $2230$  and  $1689\text{cm}^{-1}$  respectively. In case of non-functionalized MCM-41 (b) & (c) a synthesized and calcinated MCM-41 could be distinguished by non-existence of C-H stretching ( $2954$  and  $2854\text{cm}^{-1}$ ) and deformation vibrations at  $1452\text{cm}^{-1}$

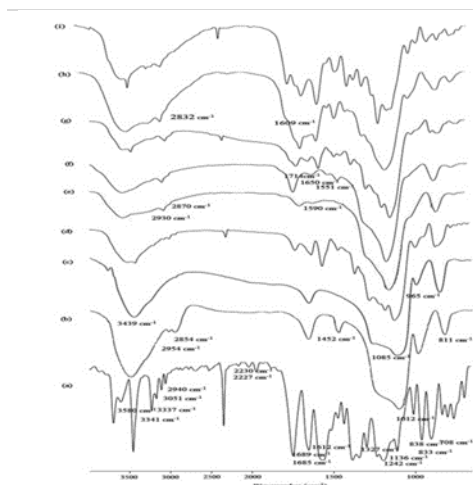


Figure.1.FT-IR Spectra (a) BIC (b) Asynthesised MSN (c) MCM-41 (d) BIC-MCM-41 (e) MCM-41-A (f) PAA-MSN (g) BIC-PAA-MSN (h) FA-MSN (i) BIC-FA-MSN

#### Differential scanning calorimetry (DSC) analysis

DSC was used in determining the melting point of the drug BIC and investigate complete entrapment at a preliminary level. The melting point of BIC was found to be 192.54 °C denoted by a sharp endothermic peak (Figure 2). The complete entrapment of drug into mesoporous carriers was confirmed by absence of any such peak in the thermogram.

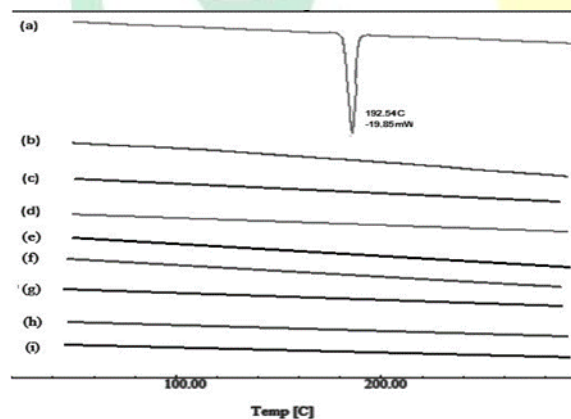


Figure.2. DSC thermograms of (a) Crystalline BIC (melting point of drug), (b) MCM-41 (c) BIC-MCM-41, (d) MCM-41-A (e) BIC-MCM-41-A (f) PAA-MSN (g) BIC-PAA-MSN (h) FA-MSN (i) BIC-FA-MSN

#### X-Ray diffraction analysis (W-XRD)

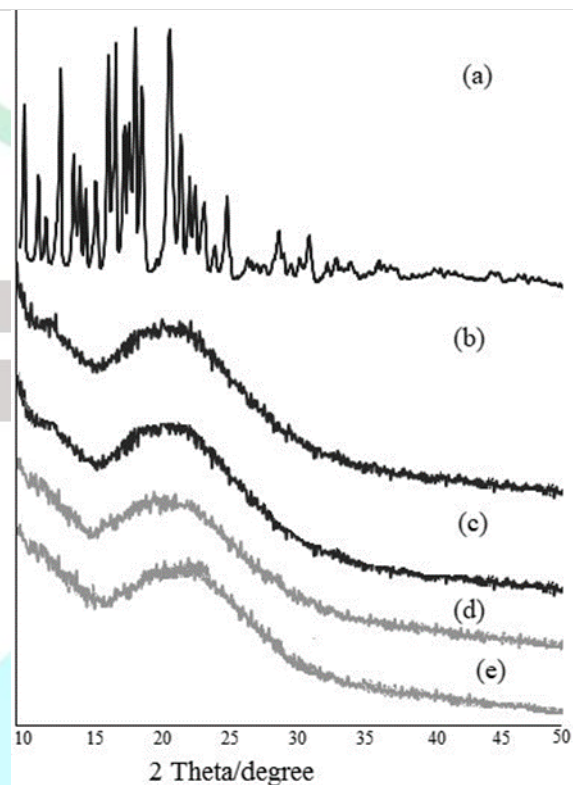


Figure.3. WAXRD spectra (a) BIC (b) BIC-MCM-41 (c) BIC-MCM-41-A (d) BIC-PAA-MSN (e) BIC-FA-MSN

Crystalline nature of BIC was justified by the sharp and highly intensified peaks obtained in the wide angle XRD spectra (Figure 3). The position of the peaks identified in order of their ascending theta values are 12.29, 18.15, 23.29, 24.11, 29.1, 29.30, 31.45, and 34.82°.

When we see the Wide-angle spectra of drug loaded MSNs, no clear intense peak is visible. This indicates the conversion of crystalline BIC to amorphous form after entrapment into MSNs. This fact could play a major part in dissolution enhancement of drug. The spectra of drug loaded bare and functionalized MSN indicated a complete entrapment of drug in to the mesopores.

#### Estimation of drug loading efficiency

The pore size and pore volume also contribute in determining the maximum amount of drug which can be entrapped in MSNs. A slight reduction in loading percentage from MCM-41 to postfunctionalization in MCM-41-A was observed.

Additionally, the TGA data also prove the thermal stability of silica materials for proposed potential application assisted directing carrier for BIC. The TGA data exhibited extent of grafting as 4% increase of MCM-41-A, 20.19% in PAA-MSNs and 23.75% increase of FA-MSNs

#### Invitro dissolution study

Drug release study was carried out in 1000 mL water with 0.5% SLS to determine the release pattern by calculating the cumulative drug release at different time points. BIC release from MCM-41 was highest followed by marketed formulation. However, a controlled and sustained behaviour of drug release was obtained in case of amine functionalized MSNs. Additionally, fast and fed state gastric and intestinal media were used to study if there is any effect of food on drug release from all the formulations (7). The drug release was almost similar in case of both fast and fed state conditions and hence it could be concluded that presence of food did not alter drug release (Fig 7.7). Thus, the medication could be taken either ways empty or after meal, it wouldn't affect the absorption or release. Further, various dissolution kinetic models were applied on the obtained % CDR data. Of the various models applied the highest regression coefficient and MSC value along with lowest AIC criteria was best fit in Weibull model in case of MCM-41 type mesoporous matrix (8) (Table 7.6 and 7.7). Whereas, in case of MCM-41-Amatrix Higuchi model was found to be the best fit satisfying all the three criteria

**In vitro diffusion study** The pH responsive based release was investigated for BIC, BIC-MCM-41, BIC-MCM-41-A, BIC-FA-MSN and BIC-PAA-MSNs at PBS media of different pH viz, 5.6, 6.8 and 7.4 as a function of time (Fig 7.8). It was observed that BIC-PAA-MSNs exhibited a highly pH responsive behavior. Drug release from PAA-MSNs was found to vary inversely and decreased with increase in pH. Maximum BIC release at 72 h was observed at pH 5.6 with percentage cumulative release being  $89.92 \pm 0.65\%$ . The release at pH 6.8 and 7.4 was found to be  $68.85 \pm 0.98\%$  and  $30.72 \pm 0.72\%$  respectively. At lower pH PAA gets completely protonated leading to enhanced release of BIC due to weakened interactions. However, for BIC-MCM-41NPs the release reached  $98.72 \pm 0.82\%$  at pH 5.6 within 48 h. Though, the release was up to 1.23 times faster in case of drug loaded bare silica when compared to BIC-PAA-MSN, no pH differentiating effect was seen, under stable BIC absence of any pH responsiveness on bare MCM-41. Further, PAA coating could also be held responsible for the slow release of BIC. A slight pH responsive behaviour was seen in case of BIC-MCM-41-A and BIC-FA-MSN but not as major as PAA-MSN. The release of BIC from PAA-MSN and MCM-41-A was faster than that of PAA-MSN. However, at higher pH like 7.4 it might be

**Conclusion:** The results obtained were suggestive of successful BIC encapsulation into the MSN shell BIC on both bare and surface functionalised. A significant enhancement in its dissolution rate and bioavailability with 3.12 and 2.61 times respectively as compared to BIC

was obtained for BIC-MCM-41. Whereas the results obtained were 1.86 and 1.38 respectively for BIC-MCM-41-A. This could aid in dose reduction and enhanced efficacy at the same time overcoming the solubility limitations. The increment in dissolution and bioavailability could also lead to dose reduction. The permeability of BIC-MCM-41NPs and BIC-MCM-41-ANPs were enhanced 4.66 and 2.71 times respectively with respect to BIC. The apoptotic assay revealed that BIC-PAA-MSN and BIC-FA-MSN were able to induce a higher

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